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REMBRANDT®

In situ Hybridisation and Detection

DISH & HRP Detection Kit-v5

DISH-HRP kit for the detection of	**Biotin Label** product code	**Digoxigenin label** product code	**# Assays**
HPV screening	A100K.0101	A100K.9901	40
HPV typing	A103K.0101	A103K.9901	40
CMV virus	A200K.0101	A200K.9901	40
Epstein-Barr virus	A300K.0101	A300K.9901	40
Herpes simplex virus	A400K.0101	A400K.9901	40

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HANDLEIDING-v4 DISH-HRP

VIAL-LABEL

METODICA D'USO-v4 DISH-HRP

SNIJDEVEN PLAKKEN VANPARAFFINE COUPES

1. Snij 4-6 µm coupes en plak ze op voorbehandelde objectglaasjes
2. Verwarm de glaasjes
3. Deparaffineer in verse xylol
4. Spoel in 100% ethanol en laat de glaasjes luchtdrogen

INCUBATIE TIJDEN

2-16 uur bij 56-60°C
2 x 10 min.
5 min.

PROTEOLYTISCHE VOORBEHANDELING

1. Los het pepsine poeder (zwart) in 4 mL gedestilleerd/gedeioniseerd water op, verdeel in 150 µL porties en bewaar bij -20°C.
2. Verdun het 1N HCl pepsine oplosmiddel (transparant) tot de concentratie die voor de applicatie nodig is (paraffine, cytologie of vries coupe; zie 3)
3. Verdun de proteolytische stock oplossing in het verdunde HCl en incubeer elk preparaat met 300-400 µL:
   paraffine 100x in 0.1N HCl;
   voeg 50 µL aan 5 mL 0.1N HCl toe
   cytologie 25.000x in 0.01N HCl;
   voeg 4µL aan 100 mL 0.01N HCl toe
   vries 50.000x in 0.01N HCl;
   voeg 2µL aan 100 mL 0.01N HCl toe
4. Overmaat van de proteolytische werkoplossing weggooien (altijd vers bereiden)
5. Ontwater de preparaten in oplopende alcoholreeks en luchtdrogen

HYBRIDISATIE PROCEDURE

1. Incubeer elk preparaat met probe reagens; 1 druppel of 20 µl en dek af met dekglaasje
2. Denatureer
3. Hybridiseer
4. Verwijder dekglaasje door preparaten in TBS buffer te dompelen
5. Incubeer elk preparaat met PanWash (wit); 5-6 druppels met uitzondering van de positieve controle
6. Spoel alle preparaten in TBS buffer

DETECTIE EN TEGENKLEURINGSPROCEDURE

1. Incubeer elk preparaat met conjugaat (rood); 2-3 druppels
2. Spoel de preparaten met TBS buffer
3. Spoel de preparaten met gedestilleerd/gedeioniseerd water
4. Bereid AEC (blauw) werkoplossing volgens onderstaande tabel

Aantal coupes	Aantal druppels AEC substraat	Volume AEC buffer
1-13	4	2mL
14-26	8	4mL
27-39	12	6mL
40-52	16	8mL

5. Incubeer elk preparaat met AEC werkoplossing; 2-3 druppels en incubeer in donker
6. Spoel de preparaten met gedestilleerd/gedeioniseerd water
7. Optioneel: incubeer elk preparaat met tegenkleuring (oranje); 2-3 druppels
8. Spoel de preparaten met gedestilleerd/gedeioniseerd water
9. Dek de coupes af

PRETRATTAMENTO DELLE SEZIONI IN PARAFFINA

1. Tagliare sezioni di 4 -6µm e depositarle su vetrino trattato
2. Scaldare i vetrini
3. Togliere la paraffina usando xilolo fresco
4. Sciacquare i vetrini in etanolo 100% ed asciugare all'aria

TEMPI DI INCUBAZIONE

2-16 ore a 56-60°C
2 x 10 min.
5 min.

TRATTAMENTO PROTEOLITICO

1. Seguendo le istruzioni del kit, sciogliere la pepsina in polvere (nera) in 4 mL di acqua distillata/deionizzata, quindi preparare aliquote di 150 µL e congelare a -20°C.
2. Diluire la pepsina 1N HCl (trasparente) diluente sino a raggiungere la concentrazione richiesta dall'applicazione (Paraffinati, Citologici o Congelati; vedasi punto 3)
3. Diluire la soluzione proteolitica scongelata in HCl diluito secondo lo schema seguente (incubare ogni sezione con 300-400 µL):
   Paraffinati: 100x in 0.1N HCl;
   aggiungere 50 µL a 5 mL 0.1N HCl
   Citologici: 25'000x in 0.01N HCl;
   aggiungere 4 µL a 100 mL 0.01N HCl
   Congelati: 50'000x in 0.01N HCl;
   aggiungere 2 µL a 100 mL 0.01N HCl
4. Gettare l'eccedenza della soluzione proteolitica
5. Disidratare le sezioni in etanolo puro ed asciugare all'aria

PROCEDIMENTO DI IBRIDAZIONE

1. Aggiungere 1 goccia o 20 µl di soluzione "probe" su ogni sezione. Coprire con coprivetrino.
2. Denaturare
3. Ibridizzare
4. Togliere il coprivetrino sciacquando i vetrino in tampone TBS
5. Aggiungere 5-6 gocce di PanWash (bianco) su ogni sezione tranne sul controllo positivo
6. Lavare tutti i vetrini in tampone TBS

PROCEDIMENTO DI DETEZIONE E COLORAZIONE

1. Aggiungere ad ogni sezione 2-3 gocce di coniugato (rosso)
2. Sciacquare i vetrini in tampone TBS
3. Sciacquare i vetrini in acqua distillata/deionizzata
4. Preparare la soluzione di lavoro dell'AEC (blu) come segue:

No. di sezioni	No. di gocce di substrato AEC	Volume di tampone AEC
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

5. Aggiungere ad ogni sezione 2-3 gocce di soluzione di lavoro AEC e incubare al buio
6. Togliere l'eccesso di soluzione substrato e lavare i vetrini in acqua distillata/deionizzata
7. Opzionale: aggiungere ad ogni sezione 2 -3 gocce di "counterstain" (arancione)
8. Lavare le sezioni in acqua distillata/deionizzata
9. Preparare le sezioni per l'osservazione al microscopio

	PEPSIN POW	DIGEST
1.	Seguendo le istruzioni del kit, sciogliere la pepsina in polvere (nera) in 4 mL di acqua distillata/deionizzata, quindi preparare aliquote di 150 µL e congelare a -20°C.	
2.		Diluire la pepsina 1N HCl (trasparente) diluente sino a raggiungere la concentrazione richiesta dall'applicazione (Paraffinati, Citologici o Congelati; vedasi punto 3)
3.	Diluire la soluzione proteolitica scongelata in HCl diluito secondo lo schema seguente (incubare ogni sezione con 300-400 µL):	
	Paraffinati: 100x in 0.1N HCl;	
	aggiungere 50 µL a 5 mL 0.1N HCl	
	Citologici: 25'000x in 0.01N HCl;	
	aggiungere 4 µL a 100 mL 0.01N HCl	
	Congelati: 50'000x in 0.01N HCl;	
	aggiungere 2 µL a 100 mL 0.01N HCl	
4.	Gettare l'eccedenza della soluzione proteolitica	
5.	Disidratare le sezioni in etanolo puro ed asciugare all'aria	

	PROBE	.....
1.	Aggiungere 1 goccia o 20 µl di soluzione "probe" su ogni sezione. Coprire con coprivetrino.	
2.	Denaturare	
3.	Ibridizzare	
4.	Togliere il coprivetrino sciacquando il vetrino in tampone TBS	
5.	Aggiungere 5-6 gocce di PanWash (bianco) su ogni sezione tranne sul controllo positivo	
6.	Lavare tutti i vetrini in tampone TBS	

	CONJ	.....	HRP
1.	Aggiungere ad ogni sezione 2-3 gocce di coniugato (rosso)		
2.	Sciacquare i vetrini in tampone TBS		
3.	Sciacquare i vetrini in acqua distillata/deionizzata		
4.	Preparare la soluzione di lavoro dell'AEC (blu) come segue:		

No. di sezioni	No. di gocce di substrato AEC	Volume di tampone AEC
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

5.	Aggiungere ad ogni sezione 2-3 gocce di soluzione di lavoro AEC e incubare al buio		
6.	Togliere l'eccesso di soluzione substrato e lavare i vetrini in acqua distillata/deionizzata		
7.	Opzionale: aggiungere ad ogni sezione 2 -3 gocce di "counterstain" (arancione)	COUNT	MG
8.	Lavare le sezioni in acqua distillata/deionizzata		
9.	Preparare le sezioni per l'osservazione al microscopio		

	WASH	TBS
1.	Togliere l'eccesso di soluzione substrato e lavare i vetrini in	
2.	acqua distillata/deionizzata	
3.	Opzionale: aggiungere ad ogni sezione 2 -3 gocce di "counterstain" (arancione)	
4.	Lavare le sezioni in acqua distillata/deionizzata	
5.	Preparare le sezioni per l'osservazione al microscopio	

	PANWASH	GC%...50%
1.	Aggiungere 5-6 gocce di PanWash (bianco) su ogni sezione tranne sul controllo positivo	
2.	Lavare tutti i vetrini in tampone TBS	

	SUBS BUFF	AEC AEC
1.	Preparare la soluzione di lavoro dell'AEC (blu) come segue:	

No. di sezioni	No. di gocce di substrato AEC	Volume di tampone AEC
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

2.	Aggiungere ad ogni sezione 2-3 gocce di soluzione di lavoro AEC e incubare al buio		
3.	Togliere l'eccesso di soluzione substrato e lavare i vetrini in acqua distillata/deionizzata		
4.	Opzionale: aggiungere ad ogni sezione 2 -3 gocce di "counterstain" (arancione)	COUNT	MG
5.	Lavare le sezioni in acqua distillata/deionizzata		
6.	Preparare le sezioni per l'osservazione al microscopio		

	TBS	.....
1.	Sciacquare i vetrini in tampone TBS	
2.	Sciacquare i vetrini in acqua distillata/deionizzata	
3.	Opzionale: aggiungere ad ogni sezione 2 -3 gocce di "counterstain" (arancione)	
4.	Lavare le sezioni in acqua distillata/deionizzata	
5.	Preparare le sezioni per l'osservazione al microscopio	

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GUIDE RÉFÉRENCE-v4 DISH-HRP

PRETRAITEMENT DES SECTIONS PARAFFINEES — *TEMPS D'INCUBATION*

1. Préparez des sections de 4-6 µm et coller les en lames traitées — 2-16 hrs à 56-60°C
2. Chauffez les lames — 2 x 10 min.
3. Deparaffinez dans du xylène frais — 5 min.
4. Immergez dans de l'éthanol 100% (ou ethanol absolu) et laissez sécher à l'air

TRAITEMENT PROTEOLYTIQUE

1. Dissolvez la poudre de pepsine (fiole) dans 4 ml d'eau distillée ou deionisée, divisez la solution en aliquotes de 150 µl et congelez les (-20°C).
2. Dilluez la solution HCl 1N (solution de pepsine en tant que diluant; transparent) selon l'usage préféré (paraffiné, cytologique ou congelé)
3. Dilluez une aliquote de la solution du stock proteolytique avec la solution diluée de HCl et incubez chaque échantillon dans 300-400 µl de la manière suivante:

<u>dilution paraffinée</u>: 100x dans 0.1N HCl;
ajouter 50 µl à 5 ml de HCl 0.01N
<u>dilution cytologique</u>: 25,000x dans 0.01N HCl
ajouter 4 µl à 100 ml HCl 0.01N
<u>dilution congelées</u>: 50,000x dans 0.01N HCl;
ajouter 2 µl à 100 ml de HCl 0.01N

4. Rincez avec de l'eau distillée ou deionisée
5. Desfydratez les lames dans une série d'éthanol et laissez sécher à l'air

PROTOCOLE D'HYBRIDATION

1. Ajoutez une goutte ou 20 µl d'une solution de sonde par échantillon et ouvrez avec une lamelle
2. Dénaturez
3. Hybridez
4. Diffusez verticalement le tampon TBS sur les lamelles
5. Ajoutez 5-6 gouttes de PanWash (blanc) à chaque échantillon à l'exception du contrôle positif
6. Rincez toutes les lames avec le tampon TBS

PROTOCOLE DE DETECTION ET DE COLORATION

1. Distribuez 23 gouttes de conjugant (rouge) par spécimen
2. Immergez les lames dans le tampon TBS
3. Immergez les lames dans de l'eau distillée ou deionisée
4. Préparez la solution d'usage AEC (bleu) à partir du tableau ci- dessous:

No. Echantillon	No. gouttes de AEC substrate	Vol. de solution tampon AEC
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

5. Ajoutez 23 gouttes de la solution d'usage AEC par spécimen et incubez dans l'obscurité.
6. Rincez les lames avec de l'eau distillée ou déionisée
7. Optionnel ajoutez 2-3 gouttes de la solution colorante (orange)
8. Rincez les lames avec de l'eau distillée ou déionisée
9. Montez les sections et examinez les lames au microscope

VIAL-LABEL

DIGEST	PEPSIN POW
DIGEST	PEPSIN DIL
PROBE	.....
PROBE	+I ..... DISH
PROBE	- I ..... DISH
PANWASH	TBS / GC% ... 50%
CONJ	..... HRP
SUBS	AEC
BUFF	AEC
COUNT	MG

GUIA DE REFERENCIA-v4 DISH-HRP

PRETRATAMIENTO DE LOS CORTES DE PARAFINA — *TIEMPO DE INCUBATION*

1. Preparar cortes de 4-6 µm y depositarlos sobre portas tratados — 2-16 horas A 56-60°C
2. Calentar los portas — 2 x 10 minutos
3. Desparafinar en xileno limpio — 5 minutos
4. Sumergir los portas en etanol absoluto y secar al aire

TRATAMIENTO PROTEOLITICO

A la recepción del kit disolver la pepsina en polvo (vial negro) en 4 ml de agua destilada/desionizada, hacer alicuotas de 150 µl y congelar a -20°C.

2. Diluir el diluyente de la pepsina (vial transparente) CH 1N a la concentración requerida para la aplicación (parafina, citología o congelación; ver 3)
3. Diluir la solución proteolítica stock descongelada en CH diludo e incubar cada muestra con 300-400 µl:

<u>Parafina</u>: 100X en CH 0.1N;
añadir 50 µl a 5 ml de CH 0.1N
<u>Citología</u>: 25.000X en CH 0.01N;
añadir 4µl a 100ml de CH 0.01N
<u>Congelación</u>: 50.000X en CH 0.01N;
añadir 2 µl a 100 ml de CH 0.01N

4. Eliminar el exceso de solución proteolítica a la dilución de trabajo
5. Desfidratar los portas en soluciones alcohólicas crecientes y secar al aire

PROBE	.....
WASH	TBS
PANWASH	TBS / GC% ... 50%
CONJ	..... HRP
WASH	TBS
SUBS	AEC
BUFF	AEC
COUNT	MG

PROTOCOLO DE HIBRIDACION

1. Añadir 1 gota o 20 µl de la solución de la sonda por muestra. Cubrir con un cubre.
2. Desnaturalizar
3. Hibridizar
4. Retirar los cubres sumergiendo los portas en tampón TBS
5. Añadir 5-6 gotas de Pan Wash (vial blanco) a cada muestra excepto al control positivo
6. Lavar todos los portas en tampón TBS

PROTOCOLO DE DETECCION Y TINCION

1. Añadir 2-3 gotas del conjugado (vial rojo) a cada muestra
2. Sumergir los portas en tampón TBS
3. Sumergir los portas en agua desionizada
4. Preparar el AEC a la solución de trabajo de acuerdo con la siguiente tabla:

Muestras	Gotas de substrato AEC	Volumen de tampón AEC
1-13	4	2 ml
14-26	8	4ml
27-39	12	6ml
40-52	16	8ml

5. Añadir 2-3 gotas de AEC a la solución de trabajo a cada muestra e incubar en oscuridad
6. Eliminar el exceso de solución de substrato y lavar los portas en agua destilada o desionizada
7. Opcionalmente añadir 2-3 gotas de solución de contraste (vial naranja) a cada muestra
8. Lavar los portas en agua desionizada
9. Montar los portas para su evaluación por microscopia

Item label description	
DIGEST	PEPSIN POW
DIGEST	PEPSIN DIL
PROBE	.....[2]
PROBE	+I .....[2] DISH
PROBE	- I .....[2] DISH
PANWASH	GC% ....[2] 50%
CONJ	.....[2] HRP
SUBS	AEC
BUFF	AEC
COUNT	MG

Item (cap) colour	Item contents description	Item amount
Black vial	: Pepsin digestion reagent	1 gram
Transparent vial	: Pepsin diluent (1N HCl solution)	15 mL
Yellow/Purple vial	: Specific[1] BIO or DIG labeled DNA probe(s)	0.8 mL
Pink vial	: DISH positive control oligo probe (BIO or DIG)	0.8 mL
Green vial	: DISH negative control DNA probe(BIO or DIG)	0.8 mL
White vial	: PanWash (differentiation reagent)	2 x 15 mL
Red vial	: HRP-conjugated anti-DIG or anti-BIO	15 mL
Blue vial	: AEC substrate	2 mL
Blue vial	: AEC buffer	15 mL
Orange vial	: Methyl Green counterstain	15 mL

Chapter 1 Introduction

1.1 Intended use

REMBRANDT® has been designed for the detection of specific DNA or RNA sequences by using the *in situ* Hybridisation (ISH) technique in paraffin embedded tissue sections, cytological specimens and frozen sections.

1.2 The ISH principle

ISH enables the detection of specific DNA or RNA sequences in histological and cytological specimens, without losing the often very essential morphological details. The principle of ISH is based on a "reaction" (= hybridisation) between a specifically labelled DNA or RNA sequence (= probe) and a DNA or RNA sequence present in the sample (= target). In case of matching sequences, a hybrid between the probe and target will be formed. Non-perfect matches are washed out by the stringency wash procedure (PanWash). The formed hybrids can easily be visualised by a specific staining procedure, i.e. substrate conversion by enzyme-conjugated antibodies. This conversion, i.e. the combination of AEC and Horseradish Peroxidase (HRP) conjugated anti-DIG or anti-BIO antibodies provided with this kit, will yield a detectable and coloured precipitation. The ISH technique is highly sensitive, specific, fast and easy to perform. Moreover, no radioactivity is involved. The reagents supplied with this kit are tailored to each other and therefore, REMBRANDT® is the ultimate user-friendly tool for performing ISH.

1.3 Controls

Use of both positive and negative controls is an essential part of the routine. To ensure that the ISH procedure is performed correctly and that observed positive and/or negative staining are specific, controls should be included in each experiment. This REMBRANDT® kit includes positive and negative control probes serving as a procedure control to be used on sections from the specimen under investigation. The positive control slides contain the desired target DNA and serve as a control for the specific probe. Additional control slides and probes are available from PanPath; please contact your local supplier.

1.4 Contents of a REMBRANDT® DISH & HRP Dectection Kit

Item (cap) colour	Item contents description	Item amount
Black vial	: Pepsin digestion reagent	1 gram
Transparent vial	: Pepsin diluent (1N HCl solution)	15 mL
Yellow/Purple vial	: Specific[1] BIO or DIG labeled DNA probe(s)	0.8 mL
Pink vial	: DISH positive control oligo probe (BIO or DIG)	0.8 mL
Green vial	: DISH negative control DNA probe(BIO or DIG)	0.8 mL
White vial	: PanWash (differentiation reagent)	2 x 15 mL
Red vial	: HRP-conjugated anti-DIG or anti-BIO	15 mL
Blue vial	: AEC substrate	2 mL
Blue vial	: AEC buffer	15 mL
Orange vial	: Methyl Green counterstain	15 mL

~ Kit contents continued ~

Item label description	
WASH	TBS
SUPPORT	GL SLIDES
SUPPORT	COVERSL
CONTROL	……[2]

Item (cap) colour	Item contents description	Item amount
White pouches	: TBS buffer salt	2 pcs
White box A	: Teflon coated glass slides, double wells	50 pcs
White box B	: Coverslips	100 pcs
White box C	: Positive control slides	2 pcs

[1] For specific probe specifications see page 9

[2] Depends on Kit specification

1.5 Materials required but not included

- Xylene for dewaxing paraffin sections.
- Fixative for cytological and frozen specimens.
- Distilled or deionised water.
- 100% Ethanol.
- 95% Ethanol.
- 70% Ethanol.
- Water-based mounting medium.
- Pipettes and tips to deliver 10-1000 µL.
- Incubation oven, set at 56-60°C to bake paraffin sections.
- Heating block/slide warmer, set at 37°C.
- Surface thermometer.
- Hotplate, set at 95°C.
- Light microscope for objective 10-100x.

1.6 Storage and shelf life

- Store all reagents at 2-8°C upon receipt of the kit.
- Store the dissolved and aliquoted pepsin reagent at -20°C, stable for at least 1 year when kept frozen.
- Store the dissolved TBS buffer at 2-8°C when not in use.
- When used and stored as indicated, the kit is stable until the expiration date printed on the box.

1.7 Safety precautions

- Some reagents contain Na-azide or thimerosal (preservation) which can cause irritation when exposed to skin or mucous membranes. The concentrations of these preservations, however, are very low (< 0.1%). If reagents come into contact with skin or eyes, rinse with large volumes of clean water.
- Never pipette solutions by mouth.
- The control slide in the kit contains pathogenic material fixed in 4% para-formaldehyde making specimens non-infectious; however, we advise taking standard precaution measures for handling infectious organisms.

REFERENCE GUIDE-v4 DISH-HRP

PRETREATMENT OF PARAFFIN SECTIONS

1. Cut 4-6 µm sections and collect on treated glass slides
2. Heat slides
3. Dewax in fresh xylene
4. Soak slides in 100% ethanol and air dry

PROTEOLYTIC TREATMENT

1. Upon receipt of the kit dissolve pepsin powder (black) in 4 mL distilled/deionized water; aliquot in 150 µL batches and freeze at -20°C
2. Dilute the 1N HCl pepsin diluent (transparent) to the application required concentration (paraffin, cytological or frozen; see 3)
3. Dilute thawed proteolytic stock solution in diluted HCl and incubate each specimen with 300-400 µL:
   paraffin: 100x in 0.1N HCl;
   add 50 µL to 5 mL 0.1N HCl
   cytological: 25,000x in 0.01N HCl;
   add 4 µL to 100 mL 0.01N HCl
   frozen: 50,000x in 0.01N HCl;
   add 2 µL to 100 mL 0.01N HCl
4. Discard excess proteolytic work solution
5. Dehydrate slides in graded ethanol and air dry

HYBRIDIZATION PROCEDURE

1. Apply 1 drop or 20 µl of probe solution per specimen; cover with coverslip
2. Denature
3. Hybridize
4. Remove coverslips by soaking slides in TBS buffer
5. Apply 5-6 drops of PanWash (white) to each specimen except the positive control
6. Wash all slides in TBS buffer

DETECTION AND STAINING PROCEDURE

1. Apply 2-3 drops of the conjugate (red) to each specimen
2. Soak slides in TBS buffer
3. Soak slides in distilled/deionized water
4. Prepare AEC (blue) work solution according the following table

Number of specimens	Number of drops of AEC substrate	Vol. of AEC buffer
1-13	4	2mL
14-26	8	4mL
27-39	12	6mL
40-52	16	8mL

5. Apply 2-3 drops of AEC work solution to each specimen and incubate in dark

5.	5-15 min. on a 37°C heating block
6.	Tap off excess substrate solution and wash slides in distilled/deionized water 3 x 1 min.
7.	Optional: apply 2-3 drops of counterstain (orange) to each specimen 1 min.
8.	Wash slides in distilled/deionized water 3 x 1 min.
9.	Mount sections for microscopic evaluation

DIGEST	PEPSIN POW	1.	Pepsin Pulver (Schwarz) in 4 mL destillierten/deionisierten Wasser lösen, in 150 µL Portionen aliquotieren und bei -20°C aufbewahren.	
DIGEST	PEPSIN DIL	2.	1N HCl Pepsin -Lösungsmittel (transparent) a.d. die Konzentration verdünnen, die für die entsprechende Anwendung notwendig ist (Paraffinschnitt, Zytologisches Präparat oder Gefrierschnitt; siehe 3)	
		3.	Verdünne die aufgetaute proteolytische Lösung in verdünnten HCl; jedes Präparat mit 300-400 µL inkubieren:	
			Paraffinschnitt: 100x in 0.1N HCl;	
			50 µL proteolytische Lösung zu 5 mL 0.1N HCl geben	30 Min. bei 37°C Heizplatte
			Zytologisches Präparat: 25,000x in 0.01N HCl;	
			4µL proteolytische Lösung zu 100 mL 0.01N HCl geben	10 Min bei 37°C Heizplatte
			Gefrierschnitt: 50,000x in 0.01N HCl;	
			2 µL proteolytische Lösung zu 100 mL 0.01N HCl geben	10 Min. bei 37°C Heizplatte
		4.	Überschüssige proteolytische Lösung verwerfen	
		5.	Schnitte in Ethanolreihe entwässern und lufttrocknen	3 x 1 Min.

HYBRIDISIERUNGSPROZEDUR

PROBE	……	1.	Tropfen oder 20 µl der Sonde auf jedes Präparat geben und mit einem Deckglas abdecken 5 Min. bei 95°C Heizplatte
		2.	Denaturieren
		3.	Hybridisieren 16 Stunden bei 37°C Ofen
WASH	TBS	4.	Entfernen der Deckgläser durch Eintauchen in TBS Puffer 10 Min.
PANWASH	TBS (GC%…50%)	5.	5-6 Tropfen PanWash (Weiss) zu jedem Präparat geben (mit Ausnahme der Positivkontrolle) 15 Min. bei 37°C Heizplatte
WASH	TBS	6.	Präparate in TBS Puffer spülen 3 x 1 Min.

DETEKTIONS- UND FÄRBEPROZEDUR

KONJ	……	1.	2-3 Tropfen Konjugat (Rot) auf jedes Präparat geben 30 Min. bei 37°C Heizplatte
WASH	TBS	2.	Präparate in TBS Puffer spülen 3 x 1 Min.
		3.	Präparate mit destilliertem/deionisierten Wasser spülen 1 Min.
		4.	AEC (Blau) Gebrauchslösung nach nachfolgendem Schema vorbereiten:

PraeparateAnzahl	Tropfen der AEC Substratloesung	Vol. AEC buffer
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

SUBS	AEC
BUFF	AEC

	5.	2-3 Tropfen der AEC Gebrauchslösung zu jedem Präparat geben 5 -15 Min. bei 37°C Heizplatte
		Im dunkeln inkubieren
		In destilliertem/deionisiertem Wasser spülen 3 x 1 Min.
COUNT	MG	7. Optional: 2-3 Tropfen der Gegenfärbelösung (Orange) auf jedes Präparat geben 1 Min.
		8. In destilliertem/deionisiertem Wasser spülen 3 x 1 Min.
		9. Präparate für die mikroskopische Beurteilung abdecken

VIAL-LABEL

ANLEITUNG-v4 DISH-HRP

HERSTELLUNG VON PARAFFINSCHNITTEN

1. 4-6 µm Schnitte anfertigen und auf vorbehandelte Objektträger ziehen
2. Schnitte inkubieren
3. In Xylol entparaffinieren
4. Schnitte in 100% Ethanol entwässern und lufttrocknen

INKUBATIONSZEITEN

2-16 Stunden bei 56-60°C	
2 x 10 Min.	
5 Min.	

PROTEOLYTISCHE BEHANDLUNG

1. Pepsin Pulver (Schwarz) in 4 mL destillierten/deionisierten Wasser lösen, in 150 µL Portionen aliquotieren und bei -20°C aufbewahren.
2. 1N HCl Pepsin -Lösungsmittel (transparent) a.d. die Konzentration verdünnen, die für die entsprechende Anwendung notwendig ist (Paraffinschnitt, Zytologisches Präparat oder Gefrierschnitt; siehe 3)
3. Verdünne die aufgetaute proteolytische Lösung in verdünnten HCl; jedes Präparat mit 300-400 µL inkubieren:
   Paraffinschnitt: 100x in 0.1N HCl;
   50 µL proteolytische Lösung zu 5 mL 0.1N HCl geben
   Zytologisches Präparat: 25,000x in 0.01N HCl;
   4µL proteolytische Lösung zu 100 mL 0.01N HCl geben
   Gefrierschnitt: 50,000x in 0.01N HCl;
   2 µL proteolytische Lösung zu 100 mL 0.01N HCl geben
4. Überschüssige proteolytische Lösung verwerfen
5. Schnitte in Ethanolreihe entwässern und lufttrocknen

HYBRIDISIERUNGSPROZEDUR

1. Tropfen oder 20 µl der Sonde auf jedes Präparat geben und mit einem Deckglas abdecken
2. Denaturieren
3. Hybridisieren
4. Entfernen der Deckgläser durch Eintauchen in TBS Puffer
5. 5-6 Tropfen PanWash (Weiss) zu jedem Präparat geben (mit Ausnahme der Positivkontrolle)
6. Präparate in TBS Puffer spülen

DETEKTIONS- UND FÄRBEPROZEDUR

1. 2-3 Tropfen Konjugat (Rot) auf jedes Präparat geben
2. Präparate in TBS Puffer spülen
3. Präparate mit destilliertem/deionisierten Wasser spülen
4. AEC (Blau) Gebrauchslösung nach nachfolgendem Schema vorbereiten:

PraeparateAnzahl	Tropfen der AEC Substratloesung	Vol. AEC buffer
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

5. 2-3 Tropfen der AEC Gebrauchslösung zu jedem Präparat geben
   Im dunkeln inkubieren
   In destilliertem/deionisiertem Wasser spülen
7. Optional: 2-3 Tropfen der Gegenfärbelösung (Orange) auf jedes Präparat geben
8. In destilliertem/deionisiertem Wasser spülen
9. Präparate für die mikroskopische Beurteilung abdecken

5 -15 Min. bei 37°C Heizplatte	
3 x 1 Min.	
1 Min.	
3 x 1 Min.	

6.5 Positive staining of the negative control

Possible causes	Remedies
■ Drying out of the section.	→ Incubate in a moisturised environment.
■ Washing procedure.	→ Make sure temperature is 37 ± 2°C. → Depending on GC%, make sure correct PanWash (Differentiation reagent) is used.
■ Contamination with positive control probe or specific probe.	→ Make sure that the positive control probe is the latest to be applied to the section.

6.6 Non-specific background staining

One should always bear in mind that the staining intensity and the level of background (or non-specific) staining may depend on the type of tissue used.

Possible causes	Remedies
■ Tissue section too thick.	→ Optimal thickness of the tissue is 4-6 µm.
■ Tissue crumbled.	→ Make sure tissue is stretched completely.
■ Deparaffinization.	→ Dewax series
■ Denaturation temperature too high.	→ Make sure temperature is 95 ± 5°C.
■ Denaturation step too long.	→ Denature no longer than 5 minutes.
■ Drying out of the section.	→ Incubate all procedure steps in a moisturised environment; prevent evaporation
■ Washing temperature.	→ Make sure temperature is 37 ± 2°C.
■ Substrate incubation step too long.	→ Shorten incubation time with 5 minutes.
■ Endogenous peroxidase.	→ Inactivate endogenous peroxidase by incubating tissue sections in 3% H_2O_2/H_2O for 15 minutes at room temperature prior to the digestion step.
■ Endogenous alkaline phosphatase.	→ Inactivate endogenous alkaline phosphatase by incubating sections in substrate solution to which 4 mg of levamisole is added.

6.7 Cross Hybridisation

One should always bear in mind that there is a possibility of cross hybridisation between related subtypes and that a patient can be infected with more than one subtype of a virus.

1.8 Performance precautions

- Read all instructions before processing any assay.
- ***DO NOT*** use reagents beyond their expiry date.
- Allow all components to warm up to room temperature (20-25°C) before use.
- Homogenize probe solution before use.
- Avoid cross contamination of specimens.
- ***DO NOT*** substitute a reagent with one from another manufacturer.
- When using treated glass slides other than those provided in the kit, specimens may fall off during the procedure.
- ***DO NOT*** perform the differentiation step on specimens incubated with the positive control oligo probe (pink)!

1.9 Preparation of reagents in advance

Pepsin digestion reagent:
Dissolve the proteolytic reagent (black) in 4 mL of distilled or deionised water (upon receipt of the kit). Aliquot in portions of i.e.50 µL and store at-20°C.

Pepsin diluent:
Dilute the 1N HCl solution (transparent) to the application required concentration (paraffin sections 0,1 N; cytological and frozen preparations 0,01 N) with distilled or deionised water.

TBS buffer salt:
Dissolve 1 pouch in 1000 mL distilled or deionised water. Dissolve the salt completely and keep the buffer free from contamination.

1.10 Preparation of the proteolytic work solution

Prepare proteolytic work solution; 300 to 400 µL per section of 1 cm^2. Make fresh work solution just before use and discard non-used solution.

Paraffin sections:
dilute aliquoted proteolytic reagent 100x in 0.1N HCl, e.g. add 50 µL to 5 mL 0.1N HCl and mix.

Cytological specimens:
dilute aliquoted proteolytic reagent 25,000x in 0.01N HCl, e.g. add 4 µL to 100 mL 0.01N HCl and mix.

Frozen sections:
Dilute aliquoted proteolytic reagent 50,000x in 0.01N HCl, e.g. add 2 µL to 100 mL 0.01N HCl and mix.

Chapter 2 REMBRANDT® DISH & HRP Detection Protocol

All incubation steps should be performed in a closed incubation chamber which contains a liquid (water) creating a saturated moisturised environment. During the incubation steps, evaporation of reagents should be prevented. Once the hybridisation procedure has been started the specimen should not be allowed to dry.

2.1 Specimen collection and pre-treatment

Paraffin embedded tissue sections

A standard procedure for tissue fixation and embedding usually involves the use of formalin and paraffin. The optimal tissue block size is 0.5 cm^3. The formalin should be buffered and fixation times should (preferably) not exceed 12 hours. Excess and/or insufficient fixation may yield suboptimal morphology and target preservation. Embedding in paraffin should not exceed temperatures of 65°C.

Sample preparation: stretch 4 µm paraffin sections on distilled water of 55°C without any additives and collect sections on bio-adhesive (i.e. organosilane) coated glass slides. Bake the slides at 56°C - 60°C in a dry air oven for 2-16 hours. Slides can be used immediately or they can be stored at room temperature for up to 3 months. Prior to ISH, slides need to be dewaxed in fresh xylene for 2 x 10 minutes. Incomplete removal of formalin and/or paraffin may affect the result of the procedure. Remove the xylene by placing the slides in 100% ethanol for 5 minutes. Air dry the slides for approximately 5-10 minutes and start with proteolytic treatment.

Cytological specimens

Make sure that no multilayer of cells is formed when making a cytological specimen. A multilayer will hamper microscopic examination of the result. The specimen be processed as soon as possible after sampling.

Sample preparation: deposit cells on coated glass slides and air dry for 30 minutes. Fix the cells with a cross-linking fixative (e.g. 4% para-formaldehyde) for 10 minutes at room temperature and rinse with PBS. Dehydrate in graded ethanol series, air dry and start with proteolytic treatment.

Frozen sections

In general, small pieces of tissue (max. 1 cm^3) are snap frozen in liquid nitrogen and either stored at -70°C or used immediately. Frozen sections are more fragile than paraffin embedded tissue sections. They should be handled with care and processed as soon as possible.

Sample preparation: collect frozen sections (4 µm) on bio-adhesive (i.e. organosilane) coated glass slides and air dry for 30 minutes. Fix the sections with a cross-linking fixative (e.g. 4% para-formaldehyde) for 10 minutes at room temperature. Dehydrate in graded ethanol series, air dry and start with proteolytic treatment.

2.2 Proteolytic treatment

Place both test and control slides on a 37°C heating block or slide warmer and add 300-400 µL of a freshly prepared, pre-warmed proteolytic work solution to each specimen. Incubate at 37°C: paraffin sections for 30 minutes, cytological and frozen specimens for 10 minutes. Tap off proteolytic work solution and dehydrate the slides in graded ethanol series (70%, 95% and 100%). Duration of each soak is 1 minute. Air dry the slides and start with the hybridisation procedure. Do not treat more than 5 slides at the same time, because the temperature of the hot plate may drop dramatically, thus causing incomplete proteolysis.

6.3 Weak or no staining on a suspected positive sample

Possible causes	Remedies
■ Tissue fixation.	→ Only use buffered formalin fixative.
■ Deparaffinization.	→ Refresh dewax series.
■ Digestion.	→ Make sure correct concentration of pepsin is used. → Make sure digestion takes place at 37°C.
■ Denaturation.	→ Make sure temperature is 95 ± 5°C.
■ Interfering internal structures of probes.	→ In case of RISH procedures, warm up probe solution at 85°C for 5 min. before usage.
■ Hybridisation procedure.	→ Homogenize probe solution prior to applying probe on the section.
■ Washing temperature.	→ Make sure temperature is 37 ± 2°C. → Depending on GC%, make sure correct PanWash (Differentiation reagent) is used.
■ Detection procedure.	→ Make sure temperature is 37 ± 2°C. → Make sure to incubate in the dark.
■ Low amount of target DNA.	→ Prolong hybridisation.
■ Colour precipitate rinsed away	→ Make sure that proper rinse and mounting media are used.

6.4 Negative staining of the positive control

Possible causes	Remedies
■ Deparaffinization	→ Re-fresh dewax series.
■ Positive control specimen incubated with positive control probe washed with PanWash. (Differentiation reagent)	→ Do not use PanWash (Differentiation reagent) on positive control specimen.
■ Denaturation temperature.	→ Make sure temperature is 95 ± 5°C.
■ Interfering internal structures of probes.	→ In case of RISH procedures, warm up probe solution at 85°C for 5 min. before usage.
■ Detection procedure.	→ Make sure temperature is 37°C ± 2°C.

Chapter 6 Trouble Shooting Guide

6.1 Introduction

This Trouble Shooting Guide is intended to support you in obtaining optimal results with PanPath's REMBRANDT® *In Situ* Hybridisation and Detection kits.

In the next pages we inform you not only about possible causes and remedies for often occurring problems when performing ISH, but we also provide you with some tips given by experts on *In Situ* hybridisation that may be of help to you.

It is of course always possible that you encounter a problem which is not covered by this Trouble Shooting Guide, or that you still have doubts about your results. In such cases, please do not hesitate to contact your local supplier or PanPath directly. Since we consider your problem as our problem, we will do our utmost to find a proper solution.

6.2 No section or cells left on the slides

Possible causes	Remedies
■ Sample preparation.	→ Make sure samples are prepared according to protocol, the tissue is fixed in neutral buffered formalin and the slides are air dried well.
■ Tissue section too thin.	→ Optimal thickness of the tissue is 4-6 µm.
■ Wrong (side of) glass slide used.	→ Use only organosilane coated glass slides.
■ Pepsin concentration too high.	→ Make sure correct concentration of pepsin is used (depending on type of specimen).
■ Digestion step too long.	→ Reduce digestion time (15 minutes instead of 30 minutes) or digest at room temperature.
■ Denaturation.	→ Make sure temperature is 95 ± 5°C. → Denature no longer than 5 minutes.
■ Coverslips removed with force.	→ Make sure that slides are soaked for at least 10 minutes in PBS.

2.3 Hybridisation procedure

Denaturation and Hybridisation

Homogenize probe solutions. Apply 1 drop or 20 µl of probe solution (yellow/purple) to each specimen and the positive control specimen. Apply 1 drop or 20 µl of the negative control probe (green) to each negative procedure control specimen and apply 1 drop or 20 µl of the positive control probe (pink) to each positive procedure control specimen. Cover all specimens with a cover slip (avoid air bubbles!). Place slides on a 95°C hotplate and incubate for 5 minutes (denaturation). Work in a preset order to ensure that all slides have been incubated at 95°C for the exact same time! Do not denature more than 5 slides at the same time, because the temperature of the hot plate may drop dramatically, thus causing incomplete denaturation. Transfer slides into a moist environment and incubate for 16 hours at 37°C (during the hybridisation the minimum temperature should be room temperature and the maximum temperature should be 37°C). Best results are obtained with prolonged incubation time (16 hours).

Differentiation (stringency wash) and rinsing

– Remove coverslips by submerging the slides in TBS buffer. Soak the slides until the coverslips fall off. Rinse the slides in TBS buffer for 10 minutes. Take the slides out, wipe off excess buffer and dry the edges using a lint-free cloth. Please mind **NOT** to perform the differentiation step on specimens incubated with the positive procedure control oligo probe (pink)!

– Apply 5-6 drops of the appropriate PanWash solution (white) to each specimen (except for the positive procedure control) and transfer the slides onto a 37°C heating block or slide warmer. Incubate for 15 minutes at 37°C. Rinse all slides 3x 1 minute in TBS buffer. Wipe off excess reagent and start with the detection and staining procedure.

2.4 Detection and staining procedure

Apply 2-3 drops of HRP-conjugate (red) to each specimen and transfer slides onto a 37°C heating block or slide warmer. Incubate for 30 minutes at 37°C. Tap off excess detection reagent and rinse the slides in TBS buffer. Soak 3x 1 minute in TBS buffer, while occasionally shaking the container. Transfer the slides into a container with distilled or deionised water and soak slides for 1 additional minute.

Prepare during the last soak the AEC work solution in a disposable polypropylene tube or suitable glassware by mixing the AEC substrate with the AEC buffer (both blue) according the volumes given below. Do not make more work solution than necessary as it deteriorates within 3 hours after production. Keep the AEC work solution well protected from the light.

# specimens	# drops of AEC substrate	volume of AEC buffer
1-13	4	2 mL
14-26	8	4 mL
27-39	12	6 mL
40-52	16	8 mL

Take the slides out, wipe off excess of water and dry around the edges using a lint-free cloth. Ensure that the specimen on the slide is not disrupted. Apply 2-3 drops of AEC substrate (blue) to each specimen and transfer the slides onto a 37°C heating block or slide warmer. Incubate in the dark for 5-15 minutes at 37°C (examine the colour development every 5 minutes microscopically). Tap off

excess substrate solution and rinse the slides for 3x 1 minute in changes of distilled or deionised water. The slides are now ready to be mounted or counterstained.

2.5 Counterstain procedure

When a contrast colour is desired, the slides can be counterstained using Methyl Green (orange). Wipe off excess reagent and apply 2-3 drops of counterstain to each specimen. Incubate for 1 minute (longer incubation is possible and will yield stronger staining). Tap off excess counterstain and rinse the slides briefly in distilled or deionised water. Mount the slides by using an aqueous mounting medium. Interpret the results under the microscope.

Contents	: - clear vial, yellow cap	= BIO labelled probe; 0.8 mL (25-40 assays)
	- clear vial, purple cap	= DIG labelled probe; 0.8 mL (25-40 assays)
	- white vial, white cap	= matching PanWash; 15 mL
		R013R.0000 for probes with GC % < 50%
		R014R.0000 for probes with GC % > 50%

Format	: ready to use
Application	: colorimetric detection of respective DNA in human specimen by *in situ* hybridisation (ISH)
Purification	: by size exclusion chromatography
Detection limit	: 3-10 pg by filter hybridisation
Storage	: refrigerated (2-8 °C); do not freeze
Stability	: until expiry date printed on label
Precautions	: - homogenise solutions before use
	- avoid contact with eyes and skin; do not swallow

Chapter 5 Probe specifications

REMBRANDT® Biotin[1] and Digoxigenin[2] labelled DNA probe specifications

PRODUCT CODE	LABEL	DNA PROBE SPECIFICATIONS			
		Description	Size	Region	Vector
A100P.0100 A100P.9900	BIO DIG	Human papilloma virus HPV screening DNA probe (PROBE xxx panHPV)*	100-300 bp	mix of total genomes 7-8 Kb; containing the conserved HPV region	pBR322; 4.3 Kb and pSP; 3.0 Kb
A191P.0100 A191P.9900	BIO DIG	Human papilloma virus type 6/11 DNA probe (PROBE xxx HPV 6/11)*	100-300 bp	total genome 7.8 Kb HPV type 6 and 7.9 Kb HPV type 11	pSP; 3.0 Kb
A192P.0100 A192P.9900	BIO DIG	Human papilloma virus type 16/18 DNA probe (PROBE xxx HPV 16/18)*	100-300 bp	total genome 7.9 Kb HPV type 16 and 7.9 Kb HPV type 18	pSP; 3.0 Kb
A193P.0100 A193P.9900	BIO DIG	Human papilloma virus type 31/33 DNA probe (PROBE xxx HPV 31/33)*	100-300 bp	total genome 7.9 Kb HPV type 31 and 7.9 Kb HPV type 33	pBR322; 4.3 Kb and modified pSP ≈ 4.0 Kb
A106P.0100 A106P.9900	BIO DIG	Human papilloma virus type 6 DNA probe (PROBE xxx HPV 6)*	100-300 bp	total genome 7.8 Kb HPV type 6	pSP; 3.0 Kb
A111P.0100 A111P.9900	BIO DIG	Human papilloma virus type 11 DNA probe (PROBE xxx HPV 11)*	100-300 bp	total genome 7.9 Kb HPV type 11	pSP; 3.0 Kb
A116P.0100 A116P.9900	BIO DIG	Human papilloma virus type 16 DNA probe (PROBE xxx HPV 16)*	100-300 bp	total genome 7.9 Kb HPV type 16	pSP; 3.0 Kb
A118P.0100 A118P.9900	BIO DIG	Human papilloma virus type 18 DNA probe (PROBE xxx HPV 18)*		total genome 7.9 Kb HPV type 18	pSP; 3.0 Kb
A131P.0100 A131P.9900	BIO DIG	Human papilloma virus type 31 DNA probe (PROBE xxx HPV 31)*	100-300 bp	total genome 7.9 Kb HPV type 31	pBR322; 4.3 Kb
A133P.0100 A133P.9900	BIO DIG	Human papilloma virus type 33 DNA probe (PROBE xxx HPV 33)*	100-300 bp	total genome 7.9 Kb HPV type 33	modified pSP ≈ 4.0 Kb
A200P.0100 A200P.9900	BIO DIG	Human Cytomegalo virus DNA probe (PROBE xxx CMV)*	100-300 bp	total genome 235 Kb	
A300P.0100 A300P.9900	BIO DIG	Epstein-Barr virus DNA probe (PROBE xxx EBV)*	100-300 bp	W fragment; 4.2 Kb	pDR720; 4.0 Kb
A400P.0100 ** A400P.9900 **	BIO DIG	Herpes simplex virus type 1/2 DNA probe (PROBE xxx HSV 1/2)*	100-300 bp	pSPM; 2.6 Kb	three HSV2 Sma I fragments;total=3.0Kb
Q001P.0100 Q001P.9900	BIO DIG	Negative control probe for DNA (CONTROL – xxx DISH)*	100-300 bp		pSP; 3.0 Kb
Q151P.0100 Q151P.9900	BIO DIG	Positive control probe for DNA (CONTROL + xxx DISH)*	30-mer oligonucleotide	Mixture of six oligonucleotides complimentary to ALU repeats	

* xxx = label (BIO or DIG)

** The HSV 1/2 probe provided with this kit stains both HSV 1 and HSV 2. In order to distinguish between HSV 1 and HSV 2, a high stringency wash ([formamid] > 60%; not provided with this kit) may be applied on consecutive sections. The high stringency wash, results in a slightly weaker staining for HSV 2 and a much weaker staining for HSV 1 when compared to results obtained with the normal in this manual described procedure. Interpretation of results are solely the responsibility of the researcher.

1 The probes in this product are labelled with the Universal Linkage System (ULS™). This product or the use of this product may be covered by one or more patents of KREATECH Biotechnology BV, including, but not restricted to, the following: EP 0539466; US 5,580,990; US 5,714,327; WO 92/01699; WO 96/35696; WO 98/15564.

2 Digoxigenin (DIG) labeling and detection is protected by international patents of Roche Molecular Biochemicals. This product is supplied under a license of Roche Molecular Biochemicals. This product or the use of this product may be covered by one or more patents of Roche Molecular Biochemicals, including the following: EP patent 0324 474 (granted); U.S. patent 5.354.657 (granted).

Chapter 3 Limitations of Procedure

3.1 Limitations

- The Rembrandt DNA and RNA *in situ* Hybridisation and Detection kits are solely applicable for the detection of corresponding DNA or RNA which may be present in cell preparations (paraffin sections, frozen sections or cytological specimen).
- Appropriate medical decisions are only possible if the medical traceability is ensured. The product is intended for professional use as an aid in the diagnosis corresponding to the DNA or RNA probes as supplied with the kit.
- Either human tissue sections or human cytological preparations may be used. Samples must be fixed in buffered formalin or alcohol. Sections should be cut at 4 um thickness, glued to the glass slides with a bio-adhesive (e.g. organosilane), dried at room temperature, subsequently dried at 37 °C overnight, complete deparaffinisation in xylene and alcohol series and air dried. Cytological specimen should be prepared as required by the user, fixed with cytological fixation agent, rinsed in distilled water prior to the ISH procedure and air dried.
- Many factors can influence the performance of the ISH procedure. Failure in detection can be due to i.e. improper sampling, handling, the time lapse between tissue removal and fixation, the size of the tissue specimen in the fixation medium, the fixation time, processing fixed tissue, the thickness of the section, the bio-adhesive on the slide, deparaffinisation procedure, incubation times, proteolytic pre-treatment, detection reagents, incubation temperatures and interpretation of results.
- The performance of the ISH procedure is also affected by the sensitivity of the method and the DNA or RNA target load; in case the limit of the sensitivity is reached or when the target DNA or RNA load is too low, a false negative reaction may be the result.
- The clinical interpretation of the results should not be established on the basis of a single test result. A precise diagnosis, in fact, should take into consideration clinical history, symptoms, as well as morphological data. Negative results therefore do not rule out any possibility of a positive specimen.
- The Rembrandt test results are not to be relied on in case the sampling, sampling method, quality, sample preparation, reagents used, controls and procedure followed is not optimal.
- Therapeutic considerations based on the result of this test alone should not been taken. Positive results should be verified by other traditional diagnostic methods such as but not limited to clinical history, symptoms, as well as morphological data.
- The medical profession should be aware of risks and factors influencing the intensity, the absence or presence of probe signals which can not be foreseen when applying this test.
- The user should carefully consider the risk and use of sample material for this test in case the sample material does not contain sufficient or representative test material.
- Laboratory personnel performing the test should be knowledgeable and be able to interpret the test results.

3.2 Interpretation of the results

First, check the negative and positive controls that have been incubated with the test slides simultaneously:

- The negative control should be really negative, i.e. not show any localised colour precipitations. If the negative control could be interpreted as being positive, discard the results since no conclusions can be drawn.

- The positive control should show colour precipitations in conformity with the localisation of the target DNA or RNA. The colour should show the proper shade and must be clearly visible in the preferential cell/ tissue type and correspond to the target localisation.

In the test slides, start under low power magnification and focus on localisation and colour to see whether:
- The positivity (colour precipitation) observed is localised in the cell type preferred by the target.
- The colour has the right shade (no endogenous or formalin pigment).

Use high power magnification to see whether:
- The positive staining texture (granular, etc), demarcation and localisation are conform the positive control staining pattern.

Product in combination with other devices

The Rembrandt *in situ* Hybridisation and Detection kits are intended for stand-alone usage. The in vitro diagnostic is intended to be used in combination with standard formalin fixed, paraffin embedded tissue blocks, standard tissue freezing, tissue sectioning (microtome), standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature and incubation time control(s), and other reagents (not supplied with this reagent) and a microscope. The combination has been tested and validated. Since the standard formalin fixed, paraffin embedded tissue blocks, standard tissue freezing, tissue sectioning (microtome), standard cytological preparation methods, hot plate(s), stove(s), incubation device(s), water bath(s), temperature controls, incubation time control(s) and other not supplied reagents such as but not limited to fixation, embedding and dewaxing reagents and a microscope is not combined with the device as a product, conformity with the essential requirements is not applicable. Assay validation criteria are mentioned in *'Interpretation of the Results'* and are also depending on the target load, which may influence the validation criteria.

Specifications of the DNA probes:

	HPV[1]	CMV	HSV	EBV	Positive Control DNA	Negative Control DNA
Specificity	95%	100%	100%	100%	100%	100%
Sensitivity	85%	85%	85%	85%	95%	95%

[1] The specificity of the HPV DNA probes is 100% for the HPV species, but the different specific HPV probe sub-types may show some inter-type cross reactions.

Chapter 4 References

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